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Control Slides For: Hepatitis

Catalog# 4280

PRODUCT DESCRIPTION

The enclosed quality control slides are intended to be used to verify histological techniques and reagent reactivity. They are to be used for the qualitative purpose of determining positive or negative results, and are not intended to be used for any quantitative purpose. The first serial section within this control box is stained and provided for your reference. **Before using these slides, review the stained slide with your pathologist to ensure that this tissue source is acceptable. We will not accept a return with missing slides in the series. Newcomer Supply guarantees reactivity of these control slides for one year from the date of receipt. Revalidate after one year to verify reactivity. Store at 15-30°C in a light deprived and humidity controlled environment.**

These control slides were produced from human surgical or autopsy tissues under carefully controlled conditions. Reasonable measures are used to deliver quality control slides that are as consistent as possible. However, characteristics of quality control slides may be dissimilar due to variations in the reagents, stains, techniques, laboratory conditions, and tissue sources used. At Newcomer Supply, we use a manual method of performing immunohistochemical and histopathology stains, specifically avoiding automation, in order to provide reactive control slides for even less aggressive methods of staining that our customers may be using.

METHOD:

Fixation: 10% Neutral Buffered Formalin
Technique: Paraffin sections cut at approximately 5 microns

COMPONENTS:

Acid Alcohol, 1%, Newcomer Supply	Catalog #10011
Ethyl Alcohol, Denatured, 70%, Newcomer Supply	Catalog #10844

Additionally needed:

Potassium Permanganate, 5% Aqueous Solution
Sulfuric Acid, 3% Aqueous Solution
Oxalic Acid, 2% Aqueous Solution
Orcein Powder

Make up the following solutions:

Potassium Permanganate Solution

Potassium Permanganate, 5% Aqueous	9.5 ml
Sulfuric Acid, 3%	5 ml
Distilled water	qs to 100 ml

Orcein Solution

Orcein Powder	1 gm
Acid Alcohol, 1%	100 ml

Add Hydrochloric Acid if necessary to adjust the pH. It should be 1-2. Allow this solution to age at least 48 hours before use.

STAINING PROCEDURE, ORCEIN METHOD

1. Deparaffinize sections in 3 changes of xylene, then hydrate through graded ethyl alcohols to distilled water.
2. Oxidize with the prepared Potassium Permanganate Solution for 10 minutes. (Proper oxidation is important for reduced nuclear and cytoplasmic staining and better contrast.)
3. Place slides in 2% Oxalic Acid for 10 minutes. Sections should be colorless.
4. Wash well in tap water.
5. Stain in Orcein Solution 4 hours or more at room temperature.
6. Rinse in 70% Alcohol.
7. Differentiate with Acid Alcohol, 1%. Control differentiation by rinsing in Ethyl Alcohol, 70%.
8. Dehydrate rapidly through graded ethyl alcohols, clear in xylene or substitute, and coverslip with a compatible mounting medium.

RESULTS:

HBsAg Dark Brown

REFERENCE:

1. "*Theory and Practice of Histotechnology*", Sheehan, D., and Hrapchak, B., 2nd edition, Batelle Press, Columbus and Richland, p. 248.
2. Modifications developed by Newcomer Supply laboratory.

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