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Part 6507 December 2016

PAP GnomePen™ Liquid Blocker – Technical Memo

PRODUCTS:	EACH
PAP GnomePen [™] – Blue	Part 6507B
PAP GnomePen [™] – Green	Part 6507G
PAP GnomePen [™] – Orange	Part 65070
PAP GnomePen [™] – Purple	Part 6507P
PAP GnomePen [™] – Red	Part 6507R
PAP GnomePen [™] – Yellow	Part 6507Y

APPLICATION:

Newcomer Supply PAP GnomePen™ Liquid Blocker, when applied around tissue sections on a microscopic slide prior to staining procedures, creates a very thin visible hydrophobic film that ensures the amount of reagent needed for sufficient reaction is greatly reduced and is retained for complete tissue section coverage. The GnomePen™ has one of the thinnest PAP pen tips available and easily allows for adjacent circles, lines or any drawn pattern to be applied on a single slide.

The GnomePen™ contains a unique formulation that is water repellent, insoluble in alcohol and acetone and soluble in xylene. A variety of barrier colors are available, allowing for color coding and specimen differentiation on the same microscopic slide. Each GnomePen™ contains approximately 350 applications in a transparent container that provides an accurate visual estimate of remaining solution volume.

METHOD:

Technique: Paraffin, frozen sections and tissue culture cells

- · Manual staining for:
 - o Immunohistochemistry (IHC) procedures
 - o Immunofluorescence Assay (IFA) procedures

PROCEDURE:

- 1. Shake pen thoroughly before use.
- Practice applying the GnomePen™ liquid barrier on a test slide; push pen tip against the glass and apply a thin liquid barrier that will dry to a film. If flow doesn't instantly begin, gently squeeze the pen body.
- 3. Store GnomePen™ tightly capped; vertically with cap end up.
- 4. Paraffin Section Method:
 - a. Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with distilled water.
 - b. Remove the slide from water or buffer; blot excess solution from the slide and around tissue section with water absorbent materials. Or place long edges of the slide on absorbent material to remove excess moisture.
 - c. The liquid barrier can be drawn on a slightly damp slide.
 - d. Encircle the tissue section(s) on the slide surface with the GnomePen™ as illustrated. Take care not to touch the pen to any edge of the tissue.
 - e. See Procedure Notes #1 and #2.
- 5. Frozen Section and Tissue Culture Cells:
 - a. Encircle the frozen tissue section(s) or tissue culture cells on the slide surface at room temperature with the GnomePen™ as illustrated. Take care not to touch the pen to any edges of the tissue or culture cells.
 - b. See Procedure Notes #1 and #2.
 - c. The liquid barrier should be applied before fixation or prior to the immersion of the slide into water or buffer.
- After liquid barrier application, allow the slide to dry in a flat position for 30-60 seconds at room temperature. Proceed with staining procedure when the drawn barrier has completely dried.
 - a. See Procedure Note #3.

- 7. Drain/rinse reagents off between staining steps; blotting slide and tissue as needed to remove excess solution.
 - a. See Procedure Note #4.
- Complete staining process and coverslip with a compatible mounting medium. GnomePen[™] barrier film will not affect the coverslipping procedure.

PROCEDURE NOTES:

- Once a tissue section is touched with GnomePen™ liquid barrier it cannot be removed. The section remains useable but a slight colorization will result on the touched portion of tissue.
- If the tissue section is not completely encircled or segregated by the GnomePen[™] barrier film, reagents will not be fully retained on the tissue section and will flood out onto the slide. This may compromise complete and adequate tissue coverage by reagents.
- If GnomePen[™] barrier lines are not completely dry prior to staining, a precipitate from reaction with detection reagents may occur.
- The use of a Slide Moisture Chamber or StainTray™ (Part 68431, 6848 or 6847) is recommended for manual staining to maintain slide organization and a moist environment during the staining process.
- GnomePen[™] is insensitive to detergents (Triton X-100, Tween 20), varying pH and temperature.

REFERENCES:

- Grizzle, William, Cecil Stockard, and Paul Billings. "The Effects of Tissue Processing Variables Other Than Fixation on Histochemical Staining and Immunohistochemical Detection of Antigens." *The Journal of Histotechnology* 24.3 (2001): 213-219.
- 2. Modifications developed by Newcomer Supply Laboratory.



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