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Part 9112 Revised October 2014

Differential Stain Kit, Smears & Touch Imprints - Technical Memo

Part 9112B KIT INCLUDES: Solution A: Xanthene Stain Solution 500ml Solution B: Thiazine Stain Solution 500ml Solution C: Fixative 500ml

Individual stain solutions may be available for purchase under separate part numbers at www.newcomersupply.com.

Xylene, ACS Part 1445

For storage requirements and expiration date refer to individual bottle labels.

APPLICATION:

The Newcomer Supply Differential Stain Kit, a modification of the Wright Giemsa Stain technique, uses aqueous based stain solutions and a methanol fixative. This stain kit provides a rapid 3-step process that can be used for differential assessment of: peripheral blood smears, touch imprints, fine needle aspirations (FNA), bone marrow biopsy aspirations, as well as detecting microorganisms.

METHOD:

Solutions: All solutions are manufactured by Newcomer Supply, Inc.

All Newcomer Supply Stain Kits are designed to be used with Coplin jars filled to 40 ml following the staining procedure provided below. Some solutions in the kit may contain extra volumes.

STAINING PROCEDURE:

- Prepare within an accepted time frame, a well-made blood smear, touch imprint, FNA smear or bone marrow aspiration smear/film per your laboratories protocol, with a focus on uniform cell distribution.
- Allow slides to thoroughly air-dry prior to staining.
- Dip dried slides in Solution C: Fixative 5-10 times, one second per dip. Allow excess fixative to drain.
- Dip slides in Solution A: Xanthene Stain Solution 5 times, one second per dip. Allow excess solution to drain.
 - See Procedure Notes #1, #2 and #3
- Quickly rinse slides with distilled water.
- Dip slides in Solution B: Thiazine Stain Solution 5 times, one second per dip. Allow excess solution to drain.
- Rinse slides quickly in distilled water. 7.
- Allow slides to air-dry, then examine microscopically.
- If coverslip is preferred, allow slides to air-dry; dip dried slides in xylene and coverslip with compatible mounting medium.

RESULTS:

Pink to yellowish-red Erythrocytes: Platelets: Violet or purple granules

Granulocytes

Eosinophils:

Nucleus - Dark blue to violet Neutrophils:

> Cytoplasm - Pale pink Granules - Purple to lilac

Nucleus - Blue Cytoplasm - Blue

Granules - Red to red-orange

Nucleus - Purple or dark blue Basophils: Granules - Dark purple

RESULTS CONTINUED:

Mononuclear Cells

Lymphocytes:

Nucleus - Violet Monocytes:

Cytoplasm - Sky blue Nucleus - Violet

Cytoplasm - Dark blue

Bacteria/microorganisms: Deep blue in varying shapes

Muscle and collagen Pale Pink Nuclei Blue/violet

Cytoplasm Varying shades of light blue

PROCEDURE NOTES:

- The division of stains in this kit gives the user the advantage of varying dips in Solutions A and B to produce different degrees of shading and intensity. However; never use fewer than three dips of one full second each.
- If more intense overall stain is desired, increase the number of dips in Solutions A and B.
 - To increase eosinophilic staining; increase the number of dips in Solution A.
 - To increase basophilic staining; increase the number of dips in Solution B.
- If a paler stain is desired; decrease dips in Solutions A and B.
- If using a xylene substitute, closely follow the manufacturer's recommendations for coverslipping application.

REFERENCES:

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- Cox, Charles. "Accuracy of Intraoperative Imprint Cytology for Sentinel Lymph Node Evaluation in the Treatment of Breast Carcinoma." Cancer Cytopathology 105.1 (2005): 13-20.
- "Guidelines of the Papanicolaou Society for Fine-Needle Aspiration Procedure and Reporting." Diagnostic Cytopathology 17 (1997): 239-247.
- McPherson, Richard and Matthew Pincus. Henry's Clinical Diagnosis and Management by Laboratory Methods. 22nd ed. Philadelphia: Elsevier Saunders, 2011. 522-535.
- Thompson, Samuel Wisley, and Ronald D. Hunt. Selected Histochemical and Histopathological Methods. 2nd Springfield, IL: Thomas, 1966. 756-762.
- Modifications developed by Newcomer Supply Laboratory.