

Steiner-Chapman Modified Silver Stain Kit - Technical Memo

KIT INCLUDES:

	Part 9172A	Part 9172B
Solution A: Zinc Formalin Sensitizer	250 ml	500 ml
Solution B: Silver Nitrate 1%, Aqueous	250 ml	500 ml
Solution C: Gum Mastic 2.5%, Alcoholic	175 ml x 2	350 ml x 2
Ingredient D: Hydroquinone, Powder	5 Grams	10 Grams
Mini Sampling Spoon		

COMPLIMENTARY POSITIVE CONTROL SLIDES: Enclosed with this kit are two complimentary unstained positive control slides to be used for the initial verification of staining techniques and reagents. Verification must be documented by running one Newcomer Supply complimentary positive control slide along with your current positive control slide for the first run. Retain the second complimentary control slide for further troubleshooting, if needed.

Individual stain solutions and additional control slides may be available for purchase under separate part numbers at www.newcomersupply.com.

Additionally Needed:

Hydrochloric Acid 5%, Aqueous	Part 12086 (for acid cleaning glassware)
Xylene, ACS	Part 1445
Alcohol, Ethyl Denatured, 100%	Part 10841
Alcohol, Ethyl Denatured, 95%	Part 10842
Acidulated Water pH 4.0-4.1	Part 10013
Coplin Jar, Plastic	Part 5184 (for microwave modification)

For storage requirements and expiration date refer to individual bottle labels.

APPLICATION:

Newcomer Supply Steiner-Chapman Modified Silver Stain Kit procedure, with included microwave modification, is used for staining spirochetes, *Helicobacter pylori*, *Legionella pneumophila*, other nonfilamentous bacteria and fungus. This modified method has eliminated Uranyl Nitrate and its regulatory and disposal requirements.

METHOD:

Fixation: Formalin 10%, Phosphate Buffered (Part 1090)

- Mercury fixatives will inhibit silver staining

Technique: Paraffin sections cut at 5 microns

Solutions: All solutions are manufactured by Newcomer Supply, Inc.

All Newcomer Supply Stain Kits are designed to be used with Coplin jars filled to 40 ml following the staining procedure provided below. Some solutions in the kit may contain extra volumes.

STAINING PROCEDURE:

- All glassware/plasticware must be acid cleaned prior to use.
 - See Procedure Notes #1 and #2 (page 2).
- Preheat Solution A: Zinc Formalin Sensitizer and Solution B: Silver Nitrate 1%, Aqueous to 60°C in a water bath.
 - Skip Step #2 if using Microwave Modification.**
- Deparaffinize sections thoroughly in three changes of xylene, 3 minutes each. Hydrate through two changes each of 100% and 95% ethyl alcohols, 10 dips each. Wash well with Acidulated Water pH 4.0-4.1 (10013); two changes, 5 minutes each.
 - See Procedure Notes #3 and #4 (page 2).
- Sensitize slides in preheated Solution A: Zinc Formalin Sensitizer in a 60°C water bath for 15 minutes.

Microwave Modification: See Procedure Note #5 (page 2).

 - Place slides in a plastic Coplin jar containing Solution A: Zinc Formalin Sensitizer and microwave to 60°C.
 - Remove from microwave; continue to incubate in warm solution under fume hood for an additional 5 minutes. Agitate occasionally for even heat distribution.
 - See Procedure Note #6 (page 2).
- Rinse in several changes of Acidulated Water pH 4.0-4.1.
- Prepare Hydroquinone Working Solution; combine and mix well.
 - Ingredient D: Hydroquinone, Powder 0.5 gm
(or one rounded scoop with reusable mini sampling spoon)
 - Distilled water 25 ml
 - Save for use in Step #12.

- Place slides in preheated Solution B: Silver Nitrate 1%, Aqueous and incubate in a 60°C water bath or oven for 1 hour.

Microwave Modification:

- Place slides in a plastic Coplin jar containing Solution B: Silver Nitrate 1%, Aqueous and microwave to 60°C.
 - Remove from microwave and let stand for an additional 1 minute 30 seconds; agitate occasionally for even heat distribution.
- Rinse well in several changes of distilled water.
 - Excessive rinsing may cause nuclei to pick up silver.
 - Dip briefly in two changes each of 95% and 100% ethyl alcohols.
 - Place slides in Solution C: Gum Mastic 2.5%, Alcoholic for 5 minutes.
 - Air-dry slides for 1-5 minutes until slides are milky white.
 - Prepare fresh Reducing Solution by combining:

a. Solution C: Gum Mastic 2.5 %, Alcoholic	15 ml
b. Hydroquinone Working Solution (Step #6)	25 ml
c. <u>Filter, then add and mix well:</u>	
d. Solution B: Silver Nitrate 1%, Aqueous	0.3 ml
 - Rinse air-dried slides quickly in distilled water.
 - Place slides in fresh Reducing Solution. Incubate in a 45°C water bath for 10-30 minutes with frequent agitation; examine microscopically at 10 minutes.
 - Check staining progress at timed intervals. Tissue will turn tan in color; bacteria will be black when the tissue reaches a golden brown color.
 - Dip in warm distilled water before and after examination.

Microwave Modification: See Procedure Note #7 (page 2).

 - Heat slides in a plastic Coplin jar containing fresh Reducing Solution and microwave at 45°C for 30 seconds.
 - Remove from microwave; continue to incubate in warm solution for an additional 2 minutes.
 - Wash for 3 minutes in running tap water; rinse in distilled water.
 - Dehydrate in two changes each of 95% and 100% ethyl alcohol. Clear in three changes of xylene, 10 dips each; coverslip with compatible mounting medium.

RESULTS:

Spirochetes	Dark brown to black
<i>Helicobacter pylori</i>	Dark brown to black
<i>Legionella pneumophila</i>	Dark brown to black
Nonfilamentous bacteria and fungus	Dark brown to black
Background	Golden brown

PROCEDURE NOTES:

1. Acid clean all glassware/plasticware (12086) and rinse thoroughly in several changes of distilled water. Cleaning glassware with bleach is not equivalent to acid washing.
2. Plastic (5500), plastic-tipped (5502, 5503), or paraffin coated metal forceps must be used with any silver solution to prevent precipitation of silver salts. No metals of any kind should be in contact with any silver solution. Only glass thermometers should be used.
3. Drain staining rack/slides after each step to prevent solution carry over.
4. The use of Acidulated Water pH 4.0-4.1 rinses (Steps #3 and #5) is recommended to ensure proper tissue pH, enhanced staining results and reduced background staining.
5. The suggested microwave procedure has been tested at Newcomer Supply using an "EB Sciences", 850 watt microwave oven with temperature probe and agitation tubes. This procedure is reproducible in our laboratory. It is nonetheless a guideline and techniques should be developed for your laboratory which meet the requirements of your situation. Microwave devices should be placed in a fume hood or vented into a fume hood, according to manufacturer's instructions, to prevent exposure to chemical vapors.
6. Heating Zinc Formalin Sensitizer will result in liberation of toxic fumes. Use appropriate protective actions.
7. The Reducing Solution contains alcohol and will reduce its boiling point. Do not boil solution. Adjust microwave times and power levels accordingly.
8. The use of some xylene substitutes may result in diminished spirochete staining. If using a xylene substitute exercise caution and closely follow the manufacturer's recommendations for deparaffinization and clearing steps.

REFERENCES:

1. Carson, Freida L., and Christa Hladik. *Histotechnology: A Self-Instructional Text*. 3rd ed. Chicago, Ill.: American Society of Clinical Pathologists, 2009. 249-250.
2. Chapman, Clifford, and Lorelei Margeson. "Use of Zinc Formalin as a Sensitizer in Silver Stains for Spirochetes." *The Journal of Histotechnology* 19.2 (1996): 135-138.
3. Churukian, Charles, and Winsome Garvey. "Microwave Steiner Method for Spirochetes and Bacteria." *The Journal of Histotechnology* 13.1 (1990): 45-47.
4. Garvey, Winsome. "Some Favorite Silver Stains." *The Journal of Histotechnology* 19.3 (1996): 269-278.
5. Sheehan, Dezna C., and Barbara B. Hrapchak. *Theory and Practice of Histotechnology*. 2nd ed. St. Louis: Mosby, 1980. 241-242.
6. Steiner, Gabriel, and Grete Steiner. "New Simple Silver Stain for Demonstration of Bacteria, Spirochetes and Fungi in Sections of Paraffin Embedded Tissue Blocks." *Journal of Laboratory Clinical Medicine* 29 (1944). 868-871.
7. Modifications developed by Newcomer Supply Laboratory.